

FLAVONOIDS FROM *Calendula officinalis* FLOWERS

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Medicinal calendula (*Calendula officinalis* L., Asteraceae) is widely cultivated in the Russian Federation and is used to produce many medicines [1, 2].

The chemical composition of *C. officinalis* includes carotinoids (a leading group of biologically active compounds), flavonoids, triterpene saponins, and several associated compounds [3, 4]. However, until now standardization of the raw material and preparations of this plant has not been fully implemented.

In our opinion, this situation is to some extent due to contradictory literature data on the flavonoid composition of medicinal calendula flowers.

Thus, one reference [5] lists the dominant component as rutin; another [6], isorhamnetin diglucoside. However, the foreign literature [7] reports that calendula flowers contain isorhamnetin 3-*O*-rutinoside.

Considering the high flavonoid content in medicinal calendula flowers and their definite significance in determining the biological activity of this material, it seemed advisable to study the component composition of calendula in order to justify the standardization of flavonoids in this medicinal plant material.

We investigated flowers of medicinal calendula (Kal'ta variety, 2006) cultivated industrially at specialized facilities in Samara District SGPU Sergievskii (Antonovka), ZAO Samaralektravy, and OOO Kentavr (Berezovka).

Ground air-dried flowers of medicinal calendula (Kal'ta variety) (100.0 g) were extracted exhaustively with ethanol (70%), combining for this maceration (24 h) and subsequent thermal extraction at 85-90°C. The aqueous ethanol extracts were evaporated in vacuo to a thick residue (about 50 mL). The condensed extract was dried over silica gel (SG) L 40/100, eluted with CHCl₃, and placed on a SG layer formed in CHCl₃. The column was eluted with CHCl₃ and CHCl₃:C₂H₅OH in various proportions (97:3, 95:5, 93:7, 90:10, 88:12, 85:15, 80:20, 70:30).

Fractions with the dominant flavonoid were combined and placed on polyamide for further purification. Dry powder (extract + polyamide) was transferred to a chromatography column (10.0 g) (4.0 cm of sorbent, 5 cm diameter) with elution by water and aqueous ethanol (20%, 40, 70, 96). The separation was monitored by TLC. The column purification over polyamide produced **1** (the dominant diagnostic flavonoid).

The next set of fractions, which also contained flavonoid-type compounds, was combined, evaporated, and dried over SG L 40/100 (1.0 g). The dry powder (extract + silica gel) was transferred to a chromatography column (4.0 cm of sorbent, 5 cm diameter). The column was eluted with CHCl₃ containing various amounts of ethanol (5%, 10, 15, 20, 25, 30, 35) to produce yellow flavonoid **2**, *R_f* 0.6.

Comparison of the chromatographic behavior of the dominant flavonoid with standard (GSO) rutin (*R_f* 0.4, CHCl₃:CH₃OH:H₂O, 26:14:3) showed that the isolated compound had a different *R_f* value (about 0.5) and; therefore, was a different flavonoid.

Isolated flavonoids **1** and **2** were identified using UV, NMR, and mass spectrometry and chemical transformations.

Compound 1, yellow crystals (EtOH), C₂₈H₃₂O₁₆, mp 173-175°C, [α]_D⁵⁴⁶ +12.0° (c 0.83, water). UV spectrum (EtOH, λ_{max}, nm): 257, 268 sh; +AlCl₃: 268, 276 sh, 359, 403. PMR spectrum [200 MHz, (CD₃)₂CO + D₂O, δ, ppm, J/Hz]: 1.04 (3H, d, J = 6.0, rhamnose CH₃), 3.2-3.8 (10H, sugar part), 3.95 (3H, s, CH₃O), 4.50 (1H, d, J = 2.0, rhamnopyranose H-1''), 5.23 (1H, d, J = 7.0, glucopyranose H-1''), 6.24 (1H, d, J = 2.0, H-6), 6.50 (1H, d, J = 2.0, H-8), 6.96 (1H, d, J = 2.0, H-5'), 7.66 (1H, dd, J = 2.0 and 8.5, H-6'), 7.97 (1H, d, J = 2.0, H-2').

Compound 2, yellow crystals (EtOH), C₂₁H₂₀O₁₂, mp 227-229°C. UV spectrum (EtOH, λ_{max}, nm): 257, 267 sh, 360; CH₃ONa: 272, 327, 410; +CH₃COONa: 272, 379; +CH₃COONa + H₃BO₃: 262, 377; +AlCl₃: 275, 412; AlCl₃ + HCl: 270, 403.

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Acid hydrolysis of **1** (2% HCl, 100°C, 1 h) produced the aglycon, which was identified as isorhamnetin {C₁₆H₁₂O₇, [M]⁺ 316, λ_{max} (EtOH) 257, 268 sh, 367 nm}, and the carbohydrate part, glucose and rhamnose. Compound **1** was hydrolyzed by rhamnodiastase, which led to the conclusion that it contained the biose rutinose. UV spectra with diagnostic reagents indicated that the isorhamnetin 3-OH was glycosylated. Therefore, **1** was isorhamnetin 3-*O*-rutinoside (narcissin).

Acid hydrolysis of **2** gave the aglycon, which was identified as quercetin {C₁₅H₁₀O₇, [M]⁺ 302, λ_{max} (EtOH) 256, 267 sh, 370 nm} and glucose. Furthermore, this glycoside was cleaved by β-glucosidase, which was consistent with a β-bond to the aglycon. A bathochromic shift of the long-wavelength band in the UV spectrum of the aglycon in the presence of AlCl₃ (+60 nm), in contrast with the starting compound (+42 nm), indicated that the 3-OH group was glycosylated. Therefore, the spectral data and chemical transformations in addition to direct comparison with an authentic sample identified **2** as quercetin 3-*O*-β-D-glucopyranoside (isoquercitrin).

Compounds **1** and **2** were isolated for the first time in the Russian Federation from cultivated medicinal calendula flowers (Kal'ta variety).

REFERENCES

1. V. A. Kurkin, *Pharmacognosy: Textbook for Students of Pharmaceutical Schools* [in Russian], OOO Ofort, GOU VPO SamSMU, Samara (2004).
2. USSR State Pharmacopeia, XIth Ed., No. 2, Meditsina, Moscow (1990).
3. R. R. Ismagilov and D. A. Kostylev, *Calendula* [in Russian], Ufa (2000).
4. E. Ya. Ladygina, *Farmatsiya*, **40**, No. 4, 84 (1992).
5. E. K. Slueva, E. N. Zhukovich, L. A. Sharikova, T. F. Pribytkova, and E. B. Derevshchikova, *Farmatsiya*, **51**, No. 1, 13 (2003).
6. V. N. Abdullabekova and A. A. Tulaganov, *Khim.-Farm. Zh.*, **35**, No. 10, 25 (2001).
7. H. Wagner, *Pharmazeutische Biologie. Drogen und ihre Inhaltsstoffe*, Gustav Fischer Verlag, Stuttgart-New York (1993).